Photocross-linking and the Cleavage of DNA by Iron Complex-substituted Psoralen

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The iron complex-substituted psoralen ($\underline{1}$) having a moiety capable of photochemically cross-linking to DNA and a DNA cleaving moiety was synthesized. The photoreaction of which is noniron complex of $\underline{1}$, with Col El plasmid DNA gave the covalently interstrand cross-linked DNA which was cleaved in the presence of FeSO and dithiothreitol under air.

Bleomycin is one of the most potent antitumor antibiotic agents known, which strongly binds to DNA and causes not only single strand but also double-strand DNA clies on. It was observed that bleomycin does not show mutagenesis although most of the antitumor antibiotics that bind to DNA, such as mitomycins, actinomycins and daunomycins, show mutagenesis or carcinogenesis. The failure of bleomycin to cause mutagenesis may be a result of the killing of cells due to irreparable damage of DNA caused by highly double strand breaks. Few reports have appeared concerning the artificial double-strand DNA-cleavage agents. Dervan has described sequence specific double-strand cleavage of DNA by bis(EDTA-distamycin.Fe). We now report the synthesis and ability of DNA strand scission of a new type of DNA cleavage agent, iron complex-substituted psoralen ($\underline{1}$).

As shown Fig. 1, the molecule was designed according to the following strategy and is essentially composed of following three parts: (A) a moiety capable of photochemically cross-linking to DNA; (B) a DNA cleaving moiety; (C) a linker moiety connecting (A) and (B) having a high binding-affinity to DNA. A psoralen was used as (A), an iron-complexing group as cleaving moiety (B) which has been synthesized as a bleomycin model, and polymethylene-amides as a linker (C).

The psoralen-linked compound $(\underline{15})$ was synthesized according to Scheme 1. 8-Hydroxypsoralen $(\underline{2})$ was refluxed with bromide $(\underline{3})$ in the presence of potassium carbonate in 2-butanone for 48 h to give $\underline{4}$ (91% yield). Hydrolysis of $\underline{4}$ with trifluoroacetic acid (TFA) gave the amine $(\underline{5})$ which was treated with the thiazolidine-2-thione derivative $(\underline{6})$, prepared by condensation of 3-(N- \underline{t} -butoxycarbonyl)aminobutanoic acid and 1,3-thazolidine-2-thione with dicyclohexylcarbodiimide (DCC) in the presence of N,N'-dimethylaminopyridine (DMAP), to

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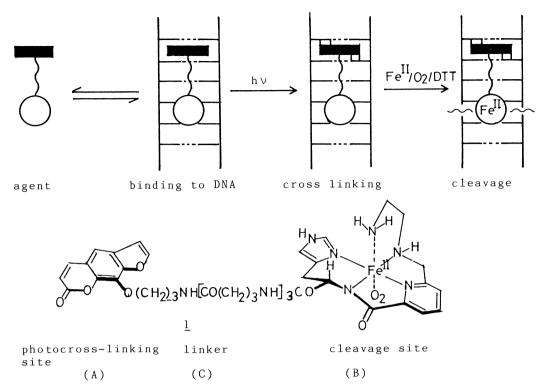


Fig. 1. Mechanism of photocross-linking and the cleavage of DNA, and structure of iron complex-substituted psoralen $(\underline{1})\,.$

give $\frac{7}{2}$ (94%). Twice repeating of the hydrolysis and amide formation of $\frac{7}{2}$ with $\frac{6}{2}$ gave $\frac{11}{2}$ (92% from $\frac{7}{2}$). Hydrolysis of $\frac{11}{2}$ with TFA gave amine $\frac{12}{2}$ which was treated with $\frac{13}{2}$ and DCC in the presence of 1-hydroxybenzotriazole (H0BT) to give $\frac{14}{8}$ (40%). Hydrolysis of $\frac{14}{2}$ with TFA in the presence of thioanisole at room temperature for 15 h gave $\frac{15}{2}$ (63%) which was purified by XAD-2 column chromatography. The UV spectum of $\frac{15}{2}$ was characteristic of psoralen at 250, 265 and 296 nm and the fast atom bombardment mass spectrum showed a molecular ion at $\frac{9}{2}$ m/z 829 (M+H) corresponding to $\frac{15}{2}$ H N O . The determination of the photosensitized cross-linking of $\frac{15}{2}$ and 8-

The determination of the photosensitized cross-linking of $\underline{15}$ and 8-methoxypsoralen $(\underline{16})$ with λDNA was performed according to the ethidium fluorescence assay. As shown in Fig. 2, $\underline{15}$ and $\underline{16}$ at 10 M concentration by irradiations were bound with covalently cross-linking to λ DNA over 80% for 30 and 60 min, respectively. The results of the photoreactions indicate that $\underline{15}$ could have a higher binding-affinity for double-stranded DNA than $\underline{16}$.

The cleavage of Co1 El supercoiled covalently closed circular (CCC) DNA(form I) with $\underline{15}$ and $\underline{16}$ to closed circular (CC) (form II) and linear (form III) DNA were carried out as follows. A solution of $\underline{15}$ or $\underline{16}$ in Tris buffer (pH 8.0) was irradiated at 360 nm at 0 °C for 45 min, and the unreacted psoralen and EDTA in the solution were removed by ultrafiltration using a microconcentrator with Tris buffer. The two cross-linked DNA preparations were reacted in the presence of FeSO, dithiothreitol (DTT) under air at 0 °C for 30 and 60 min. The reacted DNA was analyzed by means of agarose gel electrophoresis and fluorescence densitometry with ethidium bromide. The photocross-linked cccDNA (form I') of $\underline{15}$ or $\underline{16}$ moved more slowly, with tailing, than the noncross-linked form I which shows no tailing, but faster than form II and from III on the gels. As shown in Table 1, the

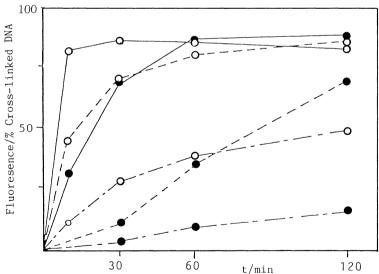


Fig. 2. Photocross-linking of $\lambda\, DNA$ with $\underline{15}$ and $\underline{16}$. Reactions were performed at 18 °C in a volume of $250\,\mu l$ buffered at pH 7.0 with 40 mM potassium phosphate, contained 24 μg of $\lambda\, DNA$, and were irradiated at 360 nm. Fluoresence reading are those after the heat denaturation and cooling cycle. O, $\underline{15}$; \bullet , $\underline{16}$; ---, $10^{-4}\, M; ----$, $10^{-5}\, M; -----$, $10^{-6}\, M$.

results of cleavages of the psoralen-cross-linked DNA were indicated that i) at < 2.5 x 10 M concentration of $\frac{15}{15}$, the presence of FeSO and DTT caused only a small increase in the cleavage (runs, 3 and 4); ii) at $> 5.0 \times 10^{-5}$ cencentrations of the substrates, the rate of (15)-cross-linked DNA cleavage increased greater than that of (16)-cross-linked DNA (runs, 6-8 and 11-13), although certain cleavage of the psoralen-cross-linked DNA was occurring during the ultrafiltration (runs, 5 and 9).

Photocross-linking and the Cleavage of DNA Using 15 and 16^{a} Table 1.

Run	Compound	b) Concentration of FeSO ₄ DTT		DTT	Time	Ratio of Forms 🔾 %		
		x 10 ⁻⁵ M	x 10 -5M	x10-3 M	min	I	ΙΙ	III
1	d)					81	19	0
2	15	1.0		1.0	30	28	67	5
3		1.0	1.0	1.0	60	25	71	4
4		2.5	1.0	1.0	60	24	67	9
5		5.0			60	46	47	7
6		5.0	1.0	1.0	30	8	82	10
7		5.0	1.0	1.0	60	0	84	16
8		5.0	5.0	1.0	60	0	80	20
9	<u>16</u>	5.0			60	52	43	5
10		5.0		1.0	60	35	61	4
11		5.0	1.0	1.0	30	35	60	5
12		5.0	1.0	1.0	60	22	73	5
13		5.0	5.0	1.0	60	14	76	10

- Photoreactions were performed with 100 W high pressure arc lamp at 360 nm 12) under argon for 45 min at 0 °C. The reaction solutions contained 6.0 µg of Col El plasmid DNA and $\frac{15}{15}$ or $\frac{16}{16}$ in a total volume of 150 µl buffered with 40 mM NaOAc /1 mM EDTA(pH 8.0). The unreacted psoralen and EDTA were removed by ultrafiltration, using centrifyed microscope trates. "CENTRICON" (AUTON) ultrafiltration using centrifugal microconcentrator "CENTRICON" (AMICON) with buffer (40 mM Tris/5 mM NaOAc)(pH 8.0). The cleavage reactions of the cross-linked DNA were performed at 0 °C in the presence or absence of FeSO₄ and /or DTT under air.
- Initial concentration of $\underline{15}$ or $\underline{16}$ in the photoreactions. Forms I, II, and III were analyzed by agarose (0.9%) gel electrophoresis and c) quantitated by densitometry after ethidium bromide staining.
- Purchased Col El plasmid DNA contained 0.6 μg.

References

- 1) L. F. Povirk, "Molecular Aspects of Anti-Cancer Drug Action," ed by S. Neidle and M. J. Waring, MacMillan Press (1983), p. 157.

- Neidle and M. J. Waring, MacMillan Press (1983), p. 157.

 2) IARC Monogr., 10, 153 (1976), ibid., 10, 29 (1976); ibid., 10, 145 (1976).

 3) P. G. Schultz and P. B. Dervan, J. Am. Chem. Soc., 105, 7748 (1983).

 4) P. -S. Song and K. J. Tapley, Jr., Photochem. Photobiol., 29, 1177 (1979).

 5) J.-P. Henichart, R. Houssin, J.-L. Bernier, and J.-P. Catteau, J. Chem. Soc., Chem. Commun., 1982, 1295; R. E. Kilkuskie, H. Suguna, B. Yellin, N. Murugesan, and S. M. Hecht, J. Am. Chem. Soc., 107, 260 (1985).

 6) E. Fujita, Pure Appl. Chem., 53, 1141 (1981).

- E. Fujita, Pure Appl. Chem., <u>53</u>, 1141 (1981).

 W. Konig and R. Geiger, Chem. Ber., <u>103</u>, 788 (1970).

 Y. Kiso, K. Ukawa, and T. Akita, J. Chem. Soc., Chem. Commun., <u>1980</u>, 101. 8)
- All compounds described gave spectral (270 MHz 1 H and 13 C 13
- 10) J. W. Lown and S.-K. Sim, Bioorg. Chem., 7, 85 (1978).
 11) J. W. Lown and A. V. Joshua, Biochem. Pharm., 29, 521 (1980); L. Giloni, M. Takeshita, F. Johnson, C. Iden, and A. P. Grollman, J. Biol. Chem., 256, 8608 (1981).
- S. T. Isaacs, C-K. J. Shen, J. E. Hearst, and H. Rapoport, Biochem., <u>16</u>, 1058 (1977).

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